Facile Synthesis of Regioselectively Deuteriated $(3\underline{R}) - (-) - [8,8,8-^2H_3] \\ \text{Linalool}$

Yoshikazu HIRAGA, Shunsuke IZUMI, Toshifumi HIRATA, and Takayuki SUGA*

Department of Chemistry, Faculty of Science, Hiroshima University,

Higashisenda-machi, Naka-ku, Hiroshima 730

Regioselectively deuteriated $(3\underline{R})-(-)-[8,8,8-^2H_3]-1$ linalool was synthesized from $(4\underline{R})-(-)-4$ -acetyl-3-methyl-5-hexenal, which was prepared from $(3\underline{R})-(-)$ -linalyl acetate, with the synthesis of $(2\underline{E},6\underline{R})-(-)-2$,6-dimethyl-2,7-[1,1- 2 H₂]octadiene-1,6-diol as the key step.

In the course of studies on the biosynthesis of cyclic monoterpenoids having a p-menthane skeleton, e.g. limonene (1), (1) we found that only $(3\underline{R})$ -isomer of linally diphosphate (LPP; (2)) was converted into $(4\underline{R})$ -(+)-limonene (1) with the enzyme preparation from Citrus unshiu. In order to examine the stereospecificity of the hydrogen elimination from the 8- or 10-position of (2) during the formation of the (3)-(0)-double bond of (1), we needed regional regional

The regioselectively deuteriated linalool (3) is synthesized from naturally occurring $(3\underline{R})$ -(-)-linally acetate (4), as shown in Scheme 1. Following the method

reported in the literature, 5) 4 (n_D^{25} 1.4473; d_4^{25} 0.8991; $[\alpha]_D^{25}$ -7.1° (neat)(lit.⁶⁾ -7.7°); 99.9% pure on GLC) was converted into (4R)-(-)-4acety1-3-methy1-5-hexenal (5) ([α] $_D^{25}$ -8.25° (c 2.0, Hexane); $\underline{m}/\underline{z}$ 94 (M⁺-AcOH)) by oxidation with m-chloroperbenzoic acid and periodic acid in 54% Wittig reaction 7) of aldehyde (5) (5 mmol) with ethyl 2triphenyl(phosphoranyidene)-propionate (6.3 mmol) in dichloromethane (15 cm³) at -20 °C gave a mixture of ethyl (2E,6R)-(-)-6-acethyl-2,6-dimethyl-2,7-octadienoate (6) and its (2Z)-isomer (95:5 on GLC) in 85% yield. mixture was subjected to column chromatography on silica gel to give the ester (6) ([α] $_{D}^{25}$ -4.4° (c 4.1, Hexane); $\underline{m}/\underline{z}$ 194 (M⁺-AcOH); IR (neat) 1730 and 1705 cm^{-1} (ester)). Reduction of 6 (2.5 mmol) with lithium aluminum deuteride (99.5% 2 H-enrichment; 6.4 mmol) in ether (10 cm 3) gave (2 \underline{E} ,6 \underline{R})- $(-)-2,6-dimethyl-2,7-[1,1-^2H_2]-octadiene-1,6-diol (7) ([\alpha]_D^{25}-16.9° (c)$ 1.18, MeOH) (lit. $^{8)}$ -12.8° for the non-deuteriated compound); 1 H NMR (270 MHz, CDCl₃) δ 1.30 (s, 3H, 10-H₃), 1.58 (m, 2H, 4-H), 1.66 (s, 3H, 9-H₃), 2.08 (m, 2H, 5-H), 5.08 (dd, 1H, J=1.5 and 10.7 Hz, 8-H(trans)), 5.22 (dd, 1H, J=1.5 and 17.6 Hz, 1-H(cis)), 5.41 (dt, 1H, J=1.5 and 7.3 Hz, 3-H), and 5.92 (dd, 1H, J=10.7 and 17.6 Hz, 7-H)] in 99% yield. spectrum of 7 showed a signal at δ 3.98 assignable to the deuterium atoms at the hydroxymethyl group. The deuterium-enrichment for 7 was found to be 99.5% by mass spectral analysis. Although it was reported that nonlabeled (2E,6R)-(-)-2,6-dimethyl-2,7-octadiene-1,6-diol could be synthe sized by the oxidation of linalool with $SeO_2^{9,10}$ or by the biological transformation of linalool with living cells, 8,11,12) these methods are recognized to give a low yield. Therefore, the method employing the Wittig reaction described above was demonstrated to be superior for the synthesis of the optically active diol.

The deuteriated diol (?) (2.0 mmol) was converted into $(2\underline{E},6\underline{R})$ -(-)-2,6-dimethyl-2,7-[1,1- 2 H $_2$]octadiene-1-chloride by treatment of \underline{N} -chlorosuccinimide (2.5 mmol) with dimethyl sulfide (0.22 mmol) in dichloromethane (15 cm 3) at -40 °C, and then the chloride was reduced with lithium aluminum deuteride (99.5% 2 H-enrichment; 5.0 mmol) to give (3 \underline{R})-(-)-3,7-dimethyl-1,6-[8,8,8- 2 H $_3$]octadien-3-ol ((3 \underline{R})-(-)-[8,8,8- 2 H $_3$]-linalool) (3) 13) (98.5% 2 H-enrichment; [α] 2_D 5 -19.1° (c 1.3, Hexane) (1it. 14) -20.1°) in 14.5% yield. A signal at δ 1.68 in the 2 H NMR spectrum of 3 indicated that the 8-methyl group of linalool was deuteriated. The overall yield of (3 \underline{R})-(-)-[8,8,8- 2 H $_3$]linalool (3) from (3 \underline{R})-(-)-linalyl acetate (4) was 7.4%.

Thus, the availability of $(3\underline{R})$ -(-)-[8,8,8- 2H_3]linalool ($\underline{3}$), which can be phosphorylated¹⁵) to yield the diphosphate ($\underline{2}$), will facilitate our studies on the stereochemistry of the biosynthesis of \underline{p} -menthane derivatives.

The authors thank Takasago Perfumery Co. Inc. for a gift of the sample of (3R)-(-)-linally acetate. The present work was in part supported by Grant-in-Aid for Scientific Research (No. 61430010) from the Ministry of Education, Science and Culture.

References

- 1) T. Suga, T. Shishibori, and H. Morinaka, J. Chem. Soc., Chem. Commun., 1980, 167.
- 2) T. Suga, T. Hirata, T. Aoki, and T. Shishibori, Phytochemistry, 25, 2769 (1986).
- 3) T. Suga, T. Hirata, S. Izumi, Y. Hiraga, and K. Okamoto, Chem. Lett., 1988, 115.
- 4) S. Izumi, Y. Katayama, Y. Hiraga, T. Hirata, and T. Suga, The 33rd Symposium on Perfume, Terpene and Essential Oil Chemistry of the Chemi-

- cal Society of Japan, Sendai, 1989, p. 107.
- 5) R. Croteau, D. M. Satterwhite, D. E. Cane, and C. C. Chang, J. Biol. Chem., 261, 13438 (1986).
- 6) A. Kaufmann and F. Kjelsberg, Am. Perfumer Essential Oil Rev., 22, 500 (1927).
- 7) R. M. Coates, D. A. Ley, and P. L. Cavender, J. Org. Chem., 43, 4915 (1978).
- 8) T. Hirata, T. Aoki, Y. Hirano, T. Ito, and T. Suga, Bull. Chem. Soc. Jpn., 54, 3527 (1981).
- 9) L. M. Stephenson and D. R. Speth, J. Org. Chem., 44, 4683 (1979).
- 10) K. B. Wiberg and S. D. Nielsen, J. Org. Chem., 29, 3353 (1964).
- 11) D. Behr, I. Wahlberg, T. Nishida, and C. R. Enzell, Acta Chem. Scand., Ser. B, 32, 228 (1978).
- 12) G. Bock, I. Benda, and P. Schreier, J. Food Sci., 51, 659 (1986).
- 13) Duteriated linalool (3): IR (neat); 3400 (OH) and 1630 cm⁻¹ (C=C); ¹H NMR (270 MHz, CDCl₃) δ 1.28 (s, 3H, 9-H₃), 1.56 (m, 2H, 4-H), 1.60 (s, 3H, 10-H₃), 2.02 (m, 2H, 5-H), 5.06 (dd, 1H, J=1.5 and 10.8 Hz, 1-H(trans)), 5.12 (dt, 1H, J=1.5 and 7.1 Hz, 6-H), 5.19 (dd, 1H, J=10.8 and 17.6 Hz, 2-H), and 5.21 (dd, 1H, J=1.5 and 17.6 Hz, 1-H(cis)); ²H NMR (76.7 MHz, CDCl₃) δ 1.68 (bs, 8-Me); ¹³C NMR (67.9 MHz, CDCl₃) δ 17.6 (C-10), 22.8 (C-5), 25.6 (bm, C-8), 27.9 (C-9), 42.1 (C-4), 73.5 (C-3), 111.7 (C-1), 124.3 (C-6), 131.9 (C-7), 145.1 (C-2); MS (rel. int.) m/z 157 (M⁺, 1%), 139 (M⁺-H₂O, 18), and 93 (100).
- 14) Y.-R. Naves, Helv. Chim. Acta, 42, 1692 (1959).
- 15) A. A. Kandutsch, H. Paulus, E. Levin, and K. Bloch, J. Biol. Chem., 239, 2508 (1964).

(Received September 27, 1990)